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Isolation and characterization of a N. CRASSA silencing gene and uses thereof

The present invention relates to the isolation and characterization of a *Neurospora crassa* gene encoding for an essential activity in the co-suppression process and to uses and applications thereof in vegetal, animal and fungine fields.

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The production of transgenic organisms is of large utility both in basic and applied biological research. The transgenic DNA is usually integrated in the genome and transferred as a Mendelian character. However, in various instances, the transgene introduction induces gene silencing phenomena (Flavell, R.B. 1994), i.e. the repression of the expression of the transgene itself and/or of one or more endogenous homologous genes.

The gene silencing (suppression of gene expression) can act two levels: transcriptional (transinactivation) where transgenes contain sequences homologous to the silenced gene promoter (Vaucheret, 1993); and post-transcriptional (co-suppression) which requires homologies between coding regions (Flavell, 1994; Stam et al., 1997; Baulcombe, 1996).

Generally the silencing induced by a transgene requires an almost complete sequence homology (from 70% to 100%) between transgene and silenced gene sequences (Elkind, 1990).

In the *Neurospora crassa* filamentous fungus, during the vegetative phase, the presence of transgenes induces a post-transcriptional gene silencing phenomenon, named "quelling" (Cogoni et al., 1996).

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By using the al-1 gene (albino 1) (Schmidhauser et al., 1990) as silencing visual marker, many features of the phenomenon have been discovered (Cogoni et al., al-1 "quelling" 1996). Particularly the gene Neurospora is characterized in that: 1) the- gene silencing is reversible further to the loss of transgene copies; 2) the reduction of mRNA basal level results from a post-transcriptional effect; 3) transgenes containing at least a region of 132 base pairs which is identical to the region encoding for the target gene are sufficient to induce the "quelling"; 4) the duplication of promoter sequences is ineffective to induce the silencing; 5) the "quelling" exhibits a dominant behavior in eterocarions containing both transgenic and untransformed nuclei, indicating the involvement of a trans-acting diffusible molecule among the nuclei; 6) the expression of an aberrant RNA transcribed by the transgenic locus is strictly correlated to silencing, suggesting that the "quelling" can be induced and/or mediated by a transgenic RNA molecule.

Therefore homologies between Neurospora silencing and plant co-suppression can be pointed out. The gene silencing in Neurospora is reversible, as result of transgenic copies instability during mitotic phase; in plants also the co-suppression reversion is associated with the reduction of transgene copy number, resulting from intra-chromosomal recombination during mitosis or meiosis (Mittelstein Scheid et al., 1994; Stam et al., 1997). Thus both in plants and in Neurospora the transgene presence is required to maintain the silencing. As in Neurospora, a decrease of the mRNA basal level of the silenced gene results from a post-transcriptional

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mechanism (Dehio and Schell 1994; van Blokand et al., 1994; de Carvalho et al., 1995). Furthermore to induce the "quelling", transgenes must contain a portion of the silencing target gene coding sequence, being the promoter region ineffective. In plants coding regions with no promoter sequences can induce silencing (van Blokand et al., 1994) and, as in the "quelling", promoters or functionally active gene products are not required for the co-suppression.

One of the similarities between "quelling" and cosuppression in plants is that both mechanisms are diffusion factors. In mediated bv eterokaryotic strains, nuclei wherein the albino-1 gene is silenced are able to induce the al-1 gene silencing of the other not transformed nuclei, all sharing the same cytoplasmic environment (Cogoni et al., 1996). In plants the presence of a diffusion factor results from the fact that the co-suppression is effective in inhibiting the replication of Tobacco Etch Virus (TEV), a RNA virus with an exclusively cytoplasmic cycle. The occurrence of highly diffusible factors, which are effective to mediate the co-suppression, has been demonstrated using the grafting technique in tobacco (Palaqui et al., 1997), showing that silenced tobacco plants are able to transfer the silencing to non-silenced plants through grafting.

The fact that "quelling" and co-suppression share all these features suggests that mechanisms involved in post-transcriptional gene silencing in plants and in fungi can be evolved by an ancestral common mechanism.

Recently gene inactivation phenomena resulting from transgene introduction have been disclosed in animals. In Drosophila melanogaster the location of a transgene close

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to heterochromatic centers results in a variegate expression (Wallrath and Elgin, 1995; Pirrotta, V., 1997). Similar expression profiles have been observed when the reference transgene is within tandem arrayed transposons, indicating that tandem repeats are effective to induce the chromatin condensation. (Dorer and Henikoff, 1994). Again in *Drosophila* Pal-Bhadra et al. (1997) have observed that the transgene introduction can lead to gene inactivation phenomena, similar to the cosuppression.

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Gene silencing phenomena resulting from transegene sequence repeats have been disclosed recently in mammalians.

Garrick et al. (1998) produced mouse transgenic lines wherein 100 transgenic copies are present in a unique locus and are repeats-arrayed in direct tandem. The transgene expression has been disclosed to be inversely proportional to the number of occurring copies, indicating that silencing phenomena dependent on repeat copies are present also in mammalians.

It has been recently found that double stranded RNA molecules can induce a sequence-specific silencing in several organisms (Fire A., 1999). The mechanism known as dsRNAi (double stranded RNA interference) acts at a post-transcriptional level by inducing sequence-specific degradation of homologous mRNAs (Montgomery, Xu and Fire, 1998). Under this aspect, dsRNAi and quelling in Neurospora are similar mechanisms, both of them acting at a post-transcriptional level. In addition, both RNA-induced silencing and DNA-induced silencing can be transmitted from cell to cell.

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Therefore the identification of *Neurospora* genes which are involved in the silencing is the first step to modulate the same process in plants, animals and fungi. The silencing modulation is of great relevance when transgenic organisms able to express the desired phenotype are produced.

The authors of the present invention have already isolated Neurospora crassa strains mutated at essential functions for gene silencing (Cogoni and Macino, 1997); 15 independent isolated mutants define three complementation groups, thus identifying the qde-1, qde-2 and qde-3 genes (qde stands for "quelling"-deficient), whose products are essential to the silencing machinery. qde genes are essential to the Neurospora silencing, as suggested by the fact that silencing of three independent genes (al-1, al-2 and qa-2) is impaired by qde mutations (Cogoni and Macino, 1997).

The authors of the present invention have already identified qde-3 gene (PCT WO 00/327885) and qde-1 gene (PCT WO 00/50581).

The authors of the invention have identified and cloned now one out of Neurospora qde genes, the qde-2 gene, thus identifying one of required factors for silencing. By considering the similarity between "quelling" and co-suppression, genes orthologous to the isolated gene are involved in co-suppression and more generally in gene silencing in other organisms, like plants, fungi and animals.

The present invention can be applied with reference to two general scopes: 1) silencing potentiation as a tool for inactivating more effectively and durably a

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desired gene, and 2) silencing suppression to obtain a better expression of the introduced transgenes.

The isolated qde-2 gene can be introduced alone or with qde-1 and/or qde-3 genes in plants, animals or fungi, in order to inactivate the expression of selected genes. The aim is to activate a sequence-specific silencing mechanism both in deficient organisms and in organisms wherein the same is not very efficient. The gene silencing can be induced also by introducing specific double stranded DNA or RNA sequences, homologous to the gene to be inactivated.

As to the silencing potentiation, the over-expression of one or more genes controlling the phenomenon can lead to higher efficiency and/or stability thereof. Therefore the introduction of qde-2 gene or of homologous genes thereof in organisms can constitute a tool to repress more effectively gene functions. Particularly this approach is specially useful in plants wherein the co-suppression is usually used for the "knock-out" of gene functions. In plants again the gene silencing potentiation can be used to obtain lines resistant to pathogen virus, by introducing transgenes encoding for viral sequences, in order to achieve the expression inhibition of the virus itself (Flavell et al., 1994).

Analogous applications are suitable for animals, wherein some indications suggest that silencing can inhibit the suitable expression of introduced transgenes (Garrick et al., 1998).

On the contrary, there are instances wherein it is desirable not to have or to reduce the gene silencing, i.e. where a transgene is to be over-expressed. It is

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known that the co-suppression is strictly correlated both with the presence of an high copy number of the transgene, and with a transgene high expression. This correlation can hamper the production of transgenic. organisms which express a transgene at high levels, because more high is the expression and/or the copy number, more probable is to evoke silencing responses. As above mentioned, analogous mechanisms inactivation, dependent on a high copy number, have been disclosed in animals. In these circumstances plant or animal lines, totally or partially ineffective for silencing, constitute an ideal recipient wherein the desired gene can be over-expressed. The invention can be applied within this scope using different approaches:

A) Identification and production of mutant lines in genes homologous to qde-2 gene, in plants, animals and fungi.

The identification of Neurospora qde-2 gene, essential for silencing mechanism, can allow the isolation of mutant lines in other organisms, mutated in genes homologous to qde-2. For example by means of amplifications using degenerated primers, designed from the most conserved regions of qde-2 gene, mutant lines in homologous genes can be identified, by analysis of insertion mutant gene banks, already available for many plant species. Both in fungi and animals such mutants can be obtained, following the identification of the homologous gene, by means of "gene disruption" techniques using homologous recombination.

B) Reduction of qde-2 gene expression

Other strategies for the production of silencingdeficient lines comprise the use of Neurospora qde-2 gene

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or homologous genes thereof. qde-2 or homologous genes can be introduced into suitable expression vectors to express them in an anti-sense orientation in order to inhibit the expression of resident endogenous genes. Alternatively portions of qde-2 or of homologous genes can be over-expressed, in order to obtain a negative dominant effect and thus blocking the function of qde-2 endogenous genes.

The authors of the present invention have cloned and characterised the *Neurospora crassa qde-2 gene*. The sequence analysis of the *qde-2* gene detected a region having a significant homology with the sequence of a *C*. elegans gene, rde-1, involved in the dsRNA mediated interference (Tabara et al., 1999).

The authors of the invention for the first time have demonstrated that the transgene induced posttranscriptional gene silencing and the dsRNA interference share common genetic mechanisms. This supports the hypothesis that the sequence specific gene silencing phenomena evolved from an ancestral mechanism aimed to protect the genome against transposons. Furthermore, the results of the authors suggest that dsRNA molecules are involved in the post-transcriptional gene silencing in fungi. dsRNA molecules could be produced directly from integrated trangenes as a result of the presence of inverted repeats or as an out come of transcription from convergent inverted promoters. Alternatively, single stranded aberrant RNA may be used as a template by an RNA-dependent RNA polymerase (such as QDE-1 protein) able to produce dsRNAs.

Within the scope of the invention the term homology is intended as similarity, i.e. number of identical

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residues + number of conserved residues with respect to the total residues of the considered sequence.

Therefore it is an object of the present invention an isolated nucleic acid molecule encoding for a protein characterized in having a silencing activity and in comprising a domain responsible for dsRNA interference, wherein the domain is at least 25% homologous with the amino acid sequence from aa. 373 to aa. 910 of sequence in fig. 1 (SEQ ID No. 2). Preferably the domain is at least 30% homologous with the amino acid sequence from aa. 373 to aa. 910 of sequence in fig. 1 (SEQ ID No. 2). More preferably the domain is at least 38% homologous with the amino acid sequence from aa. 373 to aa. 910 of sequence in fig. 1 (SEQ ID No. 2). Most preferably the domain comprises the amino acid sequence from aa. 373 to aa. 910 of sequence in fig. 1 (SEQ ID No. 2). According to a particular embodiment the isolated nucleic acid molecule encodes for a protein having the amino acid sequence of fig. 1 (SEQ ID No. 2) or functional portions thereof. Even more preferably the isolated nucleic acid molecule has the sequence of fig. 1 (SEQ ID No. 1) or its complementary sequence.

A further object of the invention is an expression vector comprising, under the control of a promoter which directs the expression in bacteria, the isolated nucleic acid molecule of the invention. Those skilled in the art will appreciate that any plasmid suitable for a correct and effective expression of the protein of the expression in bacteria can be used and it is within the scope of the invention.

A further object of the invention is an expression vector comprising, under the control of a promoter which

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directs the expression in plants or in specific plant organs, the isolated nucleic acid molecule of the invention, both in a sense and anti-sense orientation. Those skilled in the art will appreciate that any plasmid suitable for a correct and effective expression of the protein of the invention in plants or in specific plant organs can be used and it is within the scope of the invention.

A further object of the invention is an expression 10 vector comprising, under the control of a promoter which directs the expression in fungi, the isolated nucleic acid molecule of the invention, both in a sense and anti-Those skilled in the art will sense orientation. appreciate that any plasmid suitable for a correct and effective expression of the inventive protein in fungi can be used and it is within the scope of the invention.

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A further object of the invention is an expression vector comprising, under the control of a promoter which directs the expression in animals, the isolated nucleic acid molecule of the invention, both in a sense and antisense orientation. Those skilled in the art will appreciate that any plasmid suitable for a correct and effective expression of the protein of the invention in animals can be used and it is within the scope of the invention.

A further object of the invention is a prokaryotic organism transformed by using the expression vector active in bacteria of the invention.

A further object of the invention is a plant or a specific plant organ transformed by using the expression vector active in plants of the invention.

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A further object of the invention is a plant mutated at the isolated nucleic acid molecule of the invention having a reduced or inhibited silencing activity.

A further object of the invention is a fungus transformed with the expression vector of the invention active in fungi.

A further object of the invention is a fungus mutated at the isolated nucleic acid molecule of the invention and having reduced or inhibited silencing activity.

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A further object of the invention is a non-human animal transformed with the expression vector of the invention active in animals.

A further object of the invention is a non-human animal mutated at the isolated nucleic acid molecule of the invention and having a reduced or inhibited silencing activity.

A further object of the invention refers to a protein characterized in having a silencing activity and comprising а domain responsible for dsRNA interference, wherein the domain is at least 25% homologous with the amino acid sequence from aa. 373 to aa. 910 in fig. 1 (SEQ ID No. 2). Preferably the domain is at least 30% homologous with the amino acid sequence from aa. 373 to aa. 910 in fig. 1 (SEQ ID No. 2). More preferably the domain is at least 38% homologous with the amino acid sequence from aa. 373 to aa. 910 in fig. 1 (SEQ ID No. 2). Most preferably the domain comprises the amino acid sequence from aa. 373 to aa. 910 in fig. 1 (SEQ ID No. 2). According to a particular embodiment the isolated nucleic acid molecule encodes for a protein WO 01/53475

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having the amino acid sequence of fig. 1 (SEQ ID No. 2) or functional portions thereof.

It is within the scope of the present invention the use of the isolated nucleic acid molecule of the invention to modulate gene silencing in plants, animals and fungi.

The present invention now will be described by way of non limiting examples with reference to the following figures:

Figure 1: The isolated nucleic acid molecule of the 5.7 Kb fragment containing the qde-2 gene and flanking sequences (SEQ ID No.1). The amino acid sequence (SEQ ID No. 2) is shown above the nucleotide sequence.

Figure 2: It is schematically represented the pMXY2 plasmid insertion site, in the 80 mutant, used for insertional mutagenesis and consequent polimorphism of the restriction fragments by mean of DNA southern blot of a WT strain and of 80 and 820 mutant strains by using the entire restored flanking region as probe. The 820 mutant has a complete deletion of the qde-2 gene.

Figure 3: Multiple alignment, at the conserved region, among qde-2 and other proteins belonging to ago-elf2C family: A. thaliana ago-1; rabbit elf2C; C. elegans rde-1. Identical amino acids are shown in bold.

25 MATERIALS AND METHODS

E. coli strains

E. coli strain HB101 (F, hsdS20(rb, mb), supE44,
recA13, ara14, proA2, rspL20(str), xyl-5) was used for
cloning.

Neurospora crassa strains and growing conditions

Neurospora crassa following strains, supplied by Fungal Genetic Stock Center (FGSC, Dpt. Of Microbiology,

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University of Kansas Medical Ctr. Kansas City, KA) were used:

- Wild type (FGSC 987);
- qa-2/aro9 (FGSC 3957A), (FGSC 3958a).

The 6XW strain (Cogoni et al., 1996) was obtained upon transformation of the FGCS 3958a strain with pX16 plasmid (Cogoni et al., 1996). This plasmid contains the qa-2 gene used as selective marker and the al-1 coding sequence.

The mutant strains M7, M20 (qde-1); M10, M11 (qde-2); M17, M18 (qde-3) are described in Cogoni and Macino, 1997.

The qde mutants were obtained by UV mutagenesis. As recipient the transforming strain (6xw) silenced at the albino-1 gene was used. qde mutants were selected for their ability to recover a wild type unsilenced phenotype and then classified in three different complementation groups. By analyzing the al-2 gene quelling frequency all of qde used mutants are defective for the general silencing mechanism.

Complementation assays with not forced heterocaryons were carried out according to Davis and DeSerres, 1970.

Plasmids and libraries

The plasmid pMXY2, disclosed in Campbell et al.

1994, used for insertional mutagenesis was obtained from
Fungal Genetic Stock Center (FGSC, Dpt. Of Microbiology,
University of Kansas Medical Ctr. Kansas City, KA). The
plasmid contains the BmI gene (allele responsible of the
benilate drug resistance), that was used as selective
marker after transformation. The genomic DNA containing

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the qde-2 gene was isolated from a N. Crassa gene library in cosmids. (Cabibbo et al., 1991).

N. crassa transformation

Spheroplasts were prepared according to the Akins and Lambowitz (1985) protocol.

Southern Blot Analysis

Chromosomal DNA was prepared as disclosed by Irelan et al., 1993. 5 μg of genomic DNA were digested and blotted as reported in Maniatis et al.

DNA probes were: a) as to the al-1 gene the probe is represented by a XbaI-ClaI restriction fragment of pX16 (Cogoni et al., 1996); b) as to the BmI gene the probe is represented by the 2.6Kb SalI fragment of pMXY2. Northern Blot Analysis

N. crassa total RNA was extracted according to the protocol described by Cogoni et al., 1996. The mycelium was grown for two days at 30°C, then powdered in liquid nitrogen before RNA extraction. For Northern analysis 10 µg of RNA were formaldehyde denatured, electrophoresed on a 1% agarose, 7% formaldehyde gel, and blotted over Hybond N (Amersham) membranes. Hybridization was carried out in 50% formamide in the presence of ³²P labeled DNA probe 1.5x10⁶ cpm/ml.

RESULTS

25 Isolation of silencing mutant by insertional mutagenesis

Previously a Neurospora strain (6XW) wherein the albino-1 resident gene was steadily silenced was used for UV mutagenisis that brought to the isolation of qde ("quelling" deficient) mutants in $N.\ crassa$ induced gene silencing (Cogoni and Mancino 1997).

The 6XW strain shows an albino phenotype due to the lack of carotenoid biosynthesis, as results by the

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silencing of the albino 1 gene expression (Schmidhauser et al., 1990). A mutation interfering with the silencing machinery is easily detectable by producing a wild type phenotype (bright orange) of the carotenoid biosynthesis. By means of complementation assays it was possible to establish that qde mutants belong to three complementation groups, indicating the presence of three genetic loci involved in the Neurospora silencing In order to isolate the qde genes mechanism. insertional mutagenesis was carried out with the 6XW previously used for UV strain, mutagenesis. The insertional mutagenesis was carried out by transforming the 6XW strain with a plasmid, taking advantage of the fact that, after the transformation, plasmids randomly inserted in the Neurospora crassa genome. The mutagenesis was carried out transforming the 6XW silenced strain with pMXY2 (see Materials and Methods) which contains the benilate resistance as selective marker. Transformed strains able to grow in the presence of benilate containing medium and showing a wild type phenotype for the carotenoid biosynthesis were selected. Out of 50.000 isolated independent transformed strains, a benilate resistant strain (80) was isolated, which showed the bright orange phenotype expected for a qde gene mutation. In order to verify that the silencing release was effectively due to a qde gene mutation and not to the loss of al-1 transgene copies, the genomic DNA of the strain 80 was extracted and digested with SmaI and HindIII restriction enzymes. After blotting, DNA was hybridized with a probe corresponding to the coding sequence of al-1. The SmaI site is present only once in the al-1 transgene containing plasmid and the digestion

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by using said enzyme produces a 5.5Kb fragment corresponding to tandem arrayed al-1 transgenes, while a 3.1Kb fragment is expected from the resident al-1 locus. The number of al-1 transgenic copies present in the 80 strain is comparable to that present in the silenced 6XW strain.

The strain 80 is mutated in qde-2 gene

The strain 80 was assayed in a heterokaryon assay with a wild type strain and with M7, M20 (qde-1) M10, M11 (qde-2), M17, M18 (qde-3) mutants and with a wild strain (Cogoni and Macino, 1997). As shown in Table 1 the al-1 gene silencing is restored producing an albino phenotype in all of heterocaryons but M10 and M11. This behavior is consistent with the presence of a qde-2 gene recessive mutation in the strain 80.

Table 1
Reciprocal heterokaryons among the mutant 80 and previously characterized qde mutants.

	80	M7	M20	M10	M11	M17	M18
80	WT	AL	AL	WT	WT	AL	AL
M7	1	WT	WT	AL	AL	AL	AL
M20			WT	AL	AL	AL	AL
M10				WT	WT	AL	AL
M11					WT	AL	AL
M17						WT .	WT
M18							WT
			1		1		1

WT = heterokaryon with a wild type phenotype for

20 carotenoid accumulation;

AL = heterokaryon with an albino phenotype wherein the al-1 gene silencing is restored.

Recovery of sequences flanking the pMXY2 plasmid integration site

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In order to recover sequences flanking the integration site or sites the following methodology was carried out. The genomic DNA of strain 80 was digested with Aat II enzyme. Subsequently the genomic DNA was ligated and the product used to transform *E. coli* cells that was screened in an ampicillin-containing medium. PQc1 plasmid was recovered and a DNA fragment containing sequences flanking the integration site was isolated from it by using Aat II and Cla I enzymes.

Isolation of genomic clones, their subcloning and complementation of the qde-2 mutant

The fragment from pQcl plasmid was used to probe a Neurospora crassa genomic library in cosmids. Three cosmids 6G10, 20Cl and 23F2 containing about 35 Kb genomic DNA inserts, were isolated. Such cosmids were used in transformation experiments of M11 and 80 mutants. All of cosmids are able to restore the al-1 gene silencing in the two mutants, determining the appearance of an albino phenotype. The 20Cl cosmid was used to subclone a 5.7 Kb BamHI-BamHI fragment. This subclone was used for transformation experiments and resulted to be able to complement the qde-2 phenotype, indicating that a qde-2 functional gene is present in this plasmid.

Isolation and sequence of the qde-2 cDNA

The sequence of BamHI-BamHI region allowed to deduce the amino acid sequence of the QDE-2 protein. The qde-2 gene encodes for a 938 aa. putative protein (104 KDa). The genomic clone does not contain any introns since the reading frame does not contain any interruptions and intron acceptor and donor sequences were not identified (Fig. 1, Seq. ID No 1, 2).

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The qde-2 gene comprises an homologous domain with encoding genes for proteins that are responsible for dsRNA interference

The 938 aa sequence (SEQ ID No. 2) was used to search in database of amino acid sequences, by using the BLASTP algorithm. As showed in fig. 3, the search identified significant homologies with argonaute-1 gene [with expected values (E value) of 2e-57] of A. Thaliana (mutants of this gene show developmental anomalies); rde-1 gene [with expected values (E value) of 1e-23] of C. elegans, involved in gene silencing phenomena induced by double stranded RNA; elF2C gene [with expected values (E value) of 5e-60] of rabbit isolated as an element belonging to transcription beginning complex.

15 . Plant expression vector

The qde-2 gene was inserted, in a sense orientation, into a vector containing a plant expression "cassette", including the 35S promoter and the PI-II "terminator" sequences. The vector also includes the Streptomyces hygroscopicus bar gene, which confers the phosphinotricine herbicide resistance to transformed plants. In an analogous vector to the above mentioned one, qde-2 was inserted in an anti-sense orientation with respect to the 35S promoter.

The obtained vectors can be utilized to overexpress the *qde-2* gene in plants, or to repress the gene expression of resident genes, which are homologous to *qde-2*.

Fungus expression vector

The qde-2 gene was inserted in a vector containing a fungal specific expression "cassette", comprising the A. nidulans trpC gene promoter and terminator, both in a

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sense and an anti-sense orientation. In addition the vector contains the bacterial hph gene, which confers the hygromicine drug resistance. The sense plasmid can be used to over express the qde-2 gene, whereas the anti-sense plasmid is used to repress the expression of qde-2 homologous genes in various fungine species.

Mammalian expression vector

The qde-2 gene was inserted in a vector containing a mammalian specific expression "cassette", including the cytomegalovirus (CMV) promoter and SV40 termination and polyadenylation sequences both in a sense and anti-sense orientation. The vector includes also the neomicine phototransferase gene, as marker for mammalian cell selection. The sense plasmid can be used to over express the qde-2 gene, whereas the anti-sense plasmid can be used to repress the expression of qde-2 homologous genes in various mammalian species.

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Claims

- I'. An isolated nucleic acid molecule encoding for a protein characterized in having a silencing activity and in comprising a domain responsible for dsRNA interference, wherein the domain is at least 25% homologous with the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.
- 2. An isolated nucleic acid molecule encoding for a protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 1, wherein the domain is at least 30% homologous with the amino acid sequence from aa. 373 to aa. 910 of SEO ID No. 2.
 - 3. An isolated nucleic acid molecule encoding for a protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 2, wherein the domain is at least 38% homologous with the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.
 - 4. An isolated nucleic acid molecule encoding for a protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 3, wherein the domain is the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.
 - 5. An isolated nucleic acid molecule encoding for a protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 4, wherein said isolated nucleic acid molecule encodes for a protein having the amino acid sequence of SEQ ID No. 2, or functional portions thereof.

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6. An isolated nucleic acid molecule encoding for a protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 5, wherein said isolated nucleic acid

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- according to claim 5, wherein said isolated nucleic acid molecule has the sequence of SEQ ID No. 1 or its complementary sequence.
 - 7. Expression vector comprising, under the control of a promoter that directs the expression in bacteria, the isolated nucleic acid molecule according to any one of claims 1-6.
 - 8. Expression vector comprising, under the control of a promoter that directs the expression in plants or in specific plant organs, the isolated nucleic acid molecule according to any one of claims 1-6, both in a sense and anti-sense orientation.
 - 9. Expression vector comprising, under the control of a promoter that directs the expression in fungi, the isolated nucleic acid molecule according to any one of claims 1-6 both in a sense and anti-sense orientation.
 - 10. Expression vector comprising, under the control of a promoter that directs the expression in animals, the isolated nucleic acid molecule according to any one of claims 1-6 both in a sense and anti-sense orientation.
 - 11. Prokaryotic organism transformed by using the expression vector active in bacteria according to claim 7.
 - 12. Plants or a specific plant organ transformed by using the expression vector active in plants according to claim 8.
- 30 13. Plant mutated at the isolated nucleic acid molecule according to any one of claims 1-6 having a reduced or inhibited silencing activity.

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14. Fungus transformed by using the expression vector active in fungi according to claim 9.

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- 15. Fungus mutated at the isolated nucleic acid molecule according to any one of claims 1-6 having a reduced or inhibited silencing activity.
- 16. Non-human animal transformed by using the expression vector active in animals according to claim 10.
- 17. Non-human animal mutated at the isolated nucleic acid molecule according to any one of claims 1-6 having a reduced or inhibited silencing activity.
 - 18. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference wherein the domain is at least 25% homologous to the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.
 - 19. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 18 wherein the domain is at least 30% homologous to the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.
 - 20. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 19 wherein the domain is at least 38% homologous to the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.
 - 21. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 20 wherein the domain is the amino acid sequence from aa. 373 to aa. 910 of SEQ ID No. 2.

- 22. Protein characterized in having a silencing activity and comprising a domain responsible for dsRNA interference according to claim 21 comprising the amino acid sequence of SEQ ID No. 2 or functional portions thereof.
- 23. Use of the isolated nucleic acid molecule according to any one of claims 1-6 to modulate the gene silencing in plants, animals and fungi.

Length of cBANqde2.txt: 5746 bp; Listed from: 1 to: 5746; Translated from: 1039 to: 3852 (ORFs); Genetic Code used: Universal; Lun, 27 ago 1956 18:50

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69 78 87 96 105 114 123 TTT CGC TCG ATT ACT CTT TTT TTT GCG TCC GGA GTG CGA CAA AGT AGC GGC TTA TAA CAA GTC CAA 201 210 219 228 237 246 255 219 GTT GGA AAA AAA CCA TCA ATC AGT GGT ATT TCT CTC TTG GCA AAT CCA CAA CAA CCA CCC TCC ACG 267 276 285 294 303 312 321 ACA AAC AAA CAA ACA ACC TAC CTT AAC TAT CCT CTT GCT TAC CTA CGT ACC TGC CTA CCT ACC TAC 333 342 351 360 360 369 378 387 CTA CCT ACC TAC CTC TGC TGA ACC AAC CAT CTC GTC AAT CAA ACC GAA CCG AAC CAA ACC GAA CGA 399 408 417 426 426 435 444 453 TAG CCG AAT AAG CTC TCG TGC CTT GTT GCT CTC CTC GAC AAT CTG TTA CCA CCA ACA CTA CTA GTT 465 474 483 492 501 510 519 GCG GCG GAG GCC GTG GAG GTC GTG AGC GTG GAG GTG GTG GAG GCC GTG GAG GCC GTG GAG GTG GTT ACC AAG 597 606 615 624 633 642 651 GAT ACG AAC CCC CTC CAC CGG ATG TCT ACA AGT AGG TGC CTC TCC ATT TTT TAC CAT TCA ACA 888 897 TGA TGC TGA CAC GAC TTT AGG GGA ATT GAC GGT CGT GGT GCC CGC GAG CCT GAC GAC CAG ATC ACC 927 936 936 945 954 954 963 972 972 981 AAA CTC GAG GAT GAT TGG ATC AAG AAG CAC GTC AGC GAC AAT CTG GTC ACT TCC ATG AGC CAT 993 1002 1011 1020 1029 1038 1047 S L S E K E K A N N L P V R P G H G T M G E TCG CTC AGC GAG AAG GAG AAA GCC AAC AAC TTG CCG GTT CGC CCT GGC CAT GGT ACC ATG GGC GAG 1059 1068 1077 1086 1095 1104 1113 K V K L W A N Y F K I N I K S P A I Y R Y T AAG GTG AAG CTT TGG GCC AAC TAT TTC AAA ATC AAC ATC AAA TCA CCA GCC ATT TAC AGG TAC ACC ATC AAA GTT GCC GCC ACC GAG GAA AAG CTC GGA AAG GAA GCT GAG GTC GCA TCC AAG AAA GTG GAG 1209 1218 V V V G R L L K Q I E A N V K S V A I A S D GTG GTG GTT GGG AAA CTG CTC AAG CAG ATC GAA GCC AAC GTG AAA TCC GTG GCG ATT GCC AGC GAT 1275 1266 1284 1293

FIG. 1-1

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F K V H L V T T T K L K V P E N R I F E V T
TTC AAA GTG CAC CTG GTG ACG ACC ACC AAG CTC AAA GTT CCC GAG AAC CGC ATC TTT GAG GTG ACG
1323 1332 1341 1350 1359 1368 1377

W T E P S S N Q N L P S K P Q T W V V K V E TGG ACC GAG CCG AGT TCC AAC CAA AAC CTG CCC AGC CCG CCC CAG ACT TGG GTG GTC AAG GTG GAG 1389 1407 1416 1425 1425 1434 1443

E S V E T C D F G K V L N E L T T L D P K L
GAG AGT GTC GAA ACC TGC GAT TTC GGC AAG GTG CTG AAC GAG CTC ACG ACA CTT GAT CCC AAG CTC
1455 1464 1473 1482 1491 1500 1509

D G D F P K Y N V E L D A L N T I V T H H A GGG GGA GGC TTT CCC AAG TAC AAT GTG GAG CTC GAT GCC CTC AAC ACC ATT GTG ACT CAT CAT GCC 1521 1530 1539 1548 1557 1566 1575

R A D D N V A V V G R G R F F A I G D D L I CGC GCC GAC GAC AAT GTT GCG GTG GTG GGA AGG GGA AGG TTT TTT GCC ATT GGT GAT GAC CTC ATT 1587 1596 1605 1614 1623 1632 1641

E Q V R P H D S P L V I L R G Y F A S V R P GAA CAA GTG CGG CCC CAT GAC TCC CCT TTG GTC ATC TTG CGA GGA TAT TTT GCC AGC GTC CCA 1653 1662 1671 1680 1689 1698 1707

A T G R L L L N T N I T H G V F R P G V K L GCT ACC GGA AGA CTT TTA CTC AAT ACC AAC ATC ACG CAT GGT GTC TTC CGT CCT GGG GTC AAA CTT 1719 1728 1737 1746 1755 1764 1773

A Q L F Q E L G L D V M D K C N A W N E V T
GCA CAG CTG TTT CAG GAA CTT GGA CTT GAC GTA ATG GAC AAA TGC AAT GCC TGG AAC GAA GTA ACC
1785 1794 1803 1812 1821 1830 1839

K N Q L N D K M R R V H K V L A K G R V E L
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A N R G D E R G K Q K D G K E V R Y P P L F GCT AAC CGT GGC GAA GGA GGG GAG CAA AAG GAT GGT AAA GAA GTC CGA TAT CCG CCC TTG TTC 1983 1992 2001 2010 2019 2028 2037

G I P G V Q V G G P T S C Q F Y L R A R E T GGG ATT CCG GGT GTC CAG GTT GGC GGC CCG ACC TCT TGT CAG TTC TAC TTG CGT GCG CGA CAC ACA 2049 2058 2067 2076 2085 2094 2103

K D G A A P P P T P G L P S N A Y I T V A N AAG GAT GGC GCT GCC CCT CCT CCG ACT CCC GGC CTG CCG AGC AGC GCG TAC ATC ACG GTA GCG AAC 2115 2124 2133 2142 2151 2160 2169

Y Y K Q R Y G I T A N A S L P L V N V G T K
TAT TAT AAA CAA CGG TAC GGA ATA ACC GCC AAT GCT TCG CTT CCT CTG GTC AAC GTT GGC ACC AAG
2181 2190 2199 2208 2217 2226 2235

E K A I Y V L A E F C T L V K G R S V K A K GAA AAG GCC ATT TAC GTC TTG GCC GAG TTT TGT ACG CTG GTC AAA GGC CGT TCC GTC AAG GCT AAG GC

V S I O K E L I T V V G R E L K P P M L T Y
GTT TCG ATC GAC AAG GAG CTG ATC ACC GTT GTC GGG CGT GAG CTC AAG CCT CCG ATG CTT ACG TAC
2445 2454 2463 2472 2481 2490 2499

FIG. 1-2

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S G N K T V E P Q D G G W L M K F V K V A R AGC GGT AAC AAG ACG GTA GAG CCG CAG GAC GGC GGG TGG TTG ATG AAG TTT GTC AAG GTC GCC AGA 2529 2538 2547 2556 CCT TGC CGC AAG ATT GAG AAG TGG ACA TAC TTG GAA CTG AAG GGT TCC AAG GCA AAC GAA.GGG GTG 2577 2586 2595 2604 2613 2622 2631 P Q A N T A F A E F L N R T G I P I N P R F CCG CAA GCT ATG ACC GCT TTT GCC GAA TTC TTG AAC AGA ACG GGC ATC CCG ATT AAC CCC AGG TTC 2643 2652 2661 2670 2679 2688 2597 P G M S M S V P G S E K E F F A K V K TCG CCG GGC ATG AGC ATG TCA GTT CCA GGG AGC GAA AAA GAG TTC TTT GCC AAA GTG AAG GAA CTC 2709 2718 2727 2736 2745 2754 2763 2793 2802 K R A A D I T F G V H T V C C V A E K F L S AAG CGG GCT GCC GAT ATC ACA TTT GGC GTT CAC ACA GTC TGT TGT GTA GCC GAA AAG TTC CTT AGC 2841 2850 2859 2868 2877 2886 2895 T K G Q L G Y F A N V G L K V N L K F G G T ACT AAG GGG CAG CTG GGG TAT TTT GGC AAC GTC GGC CTC AAG GTC AAC CTC AAG TTT GGC GGC ACC 2907 2916 2925 2934 2943 2952 2961N H N I K T P I P L L A K G K T M V V G Y D
AAT CAC AAT ATC AAG ACG CCC ATT CCT TTG CTC GCC AAG GGG AAG ACG ATG GTG GGC TAT GAT
2973 2982 2991 3000 3009 3018 3027 V T H P T N L A A G Q S P A S A P S I V G L GCC CAT CCG ACC AAT CTA GCG GCT GGA CAA TCG CCT GCA TCG GCT CCC AGT ATT GTC GGC CTG 3039 3048 3057 3066 3075 3086 2002 A N N R S L P E N I L I F R D G V S E G Q F GCA AAC AAC CGC AGT CTC CCC GAG AAT ATC CTG ATT TTC CGC GAT GGC GTC TCC GAG GGA CAG TTC 3237 3246 3255 3264 3273 3282 3291 Q M V I K D E L P L V R A A C K L V CAG ATG GTC ATC AAG GAC GAG CTA CCC CTG GTT CGC GCC GCC TGC AAG CTG GTG TAT CCA GCT GGC 3312 3321 3330 3339 K L P R I T L I V S V K R H Q T R F F P T D
AAG CTA CCG CGT ATT ACG CTG ATT GTC TCT GTC AAG CGC CAC CAG ACT CGC TTC TTC CCA ACG GAC 3387 3396 3405 3414 CCG AAG CAT ATT CAC TTC AAG TCC AAG AGC CCC AAG GAG GGT ACT GTG GTT GAC CGC GGC GTG ACC 3444 3453 3462 N V R Y W D F F L Q A H A S L Q G T A R S A AAC GTC CGC TAT TGG GAC TTC TTT TTG CAG GCG CAC GCG TCG CTC CAG GGC ACG GCC CGC TCG GCT 3501 3510 3519 3528 3537 3546 3555H Y T V L V D E I F R A D Y G N K A A D T L CAG GTG GTG GAT GAG ATT TTC AGG GCC GAC TAT GGA AAC AAG GCG GCC GAC ACG CTG 3567 3576 3585 3594 3603 3612 3621 E Q L T H D M C Y L F G R A T K A V S I C P GAG CAG CTG ACG CAT GAC ATG TGT TAT CTC TTT GGA CGC CCC ACG GCT GTC AGT ATC TGC CCG $^{\circ}$

FIG. 1-3

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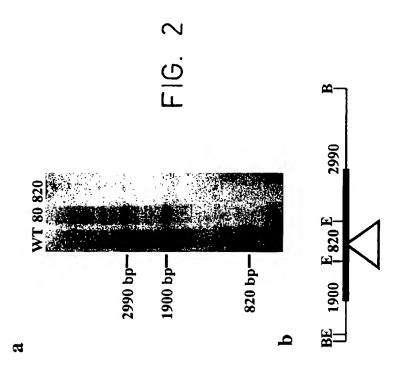
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FIG. 1-4

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	AAA GAA 5151		2160	,	2169			5178			5187		:	5196		520	5
GGA	AAA AAC 5217	GGA	GAA GGA 5226	AAA	AAA AAA 5235	CAT	AAZ	5244	AAA	AAA	5253	AAC		AAG 5262	AAA	GAA CT. 527	
AAT	CAT CCA 5283	AAC	TCA GCG 5292	GAA	AGT ACT 5301	CAT	ACA	5310	GGT	CGG	СТG 5319	CCT		TCG 328	GAC	TCC CC.	
TCT	5349	GGT	ACT GAT 5358	TCT	GCT GCC 5367	CCA	GAC	TTC 5376	CAC	TTT	CAA 5385	AGT		TAT 5394	CAC	CCT TA:	
TGT	TAG AGT 5415	GAG	TAG TAG 5424	ACG	TAA GTC 5433	CTC	CCG	ATC 5442	CGG	AGC	CAA 5451	AAC		TCC 460	CTT	TCC CAC	
TAT	CCC TCT 5481	TCA	ATC CAC 5490	CAG	TAG CAA 5499	CAC	CCA	ТСТ 5508	TGC	CAT	AGA 5517	GCG		TAT 526	ccc	CTG CCC	
CCC	CTG CCG 5547	AGC	CAG GAG 5556	TAG	CAG TCC 5565	TAT	TCA	TAG 5574	GCG	GAC	TCC 5583	TCT		CGT 592	CTT	CCG ACA	GGG
ACA	5613	TTG	GTA GGG 5622	CAC	CCG CAG 5631	CAG	AGG	AGG 5640	AGG	TAT	TTC 5649	TGT		GAC 658	TGG	TTC TGT 5667	TTG
GGG	CAG CTA 5679	AGG	GCG TGG 5688	GT T	TCC TTC 5697	GTG	AGC	CGC 5706	TGT	TGT 5	GAT 5715	TGT		CGG 724	CGG	CGT CCG 5733	AGG
ATA	AGG ATC 5745	С															

FIG. 1-5



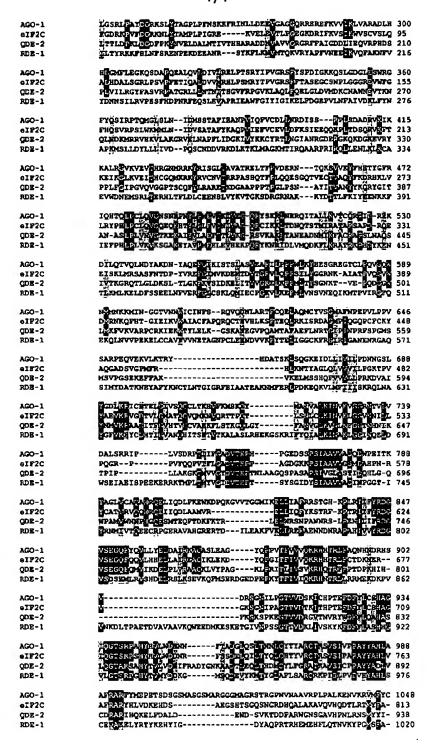


FIG. 3

SEQUENCE LISTING

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<120> Isolation and characterization of a N. crassa silencing gene and uses thereof

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<150> RM2000A000021

<151> 2000-01-17

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gaggagtagt egtttegete gattaetett tttttgegt eeggagtgeg acaaagtage 240

ggettataac aagtecaagt tggaaaaaaa eeateaatea gtggtatte tetettggea 300

aatecacaac aateceette eacgacaaac aaacaaacaa eetaeettaa etateetett 360

gettaectae gtaeetgeet acetaeetae etaeetaeet aeetetgete aaceaaceat 420

ctegteaate aaacegaace gaaceaaace gaacgatage egaataaget etegtgeett 480

gt	tgct	ctac	tcg	acaa	tct	gtta	ccac	ca a	cact	acaa	g tt	taac	agtc	atg	tctgac	a 540
at	cgtg	gcgg	tcg	tgga	ggt	cgtg	gcgg	cg g	tggt	cgcg	g cg	gcgg	cggc	9 9c	ggcgga	g 600
gc	cgtg	gagg	tgg	tçag	caa	ggcg	geggt	tg g	aggc	cgtg	g ag	gtgg	ttac	caa	ggcagc	g 660
gc	ggcg	gtgg	agg	ccgt	ggc	ggcg	gttai	tc a	aggc	ggtg	g cg	gcgg	tgac	cgt	ggaggc	c 720
gt	ggcg	gcgg	tta	tcaaq	ggc	ggtg	gtggd	g gi	tggt	ttcc	a ag	gegge	eggt	ggaa	aggggt	g 780
gco	gtg	gcgg	cgg	tttc	caa (ggcgg	gegge	g go	cggc	ggcc	g tg	gtggd	ettc	ggc	ggagga	c 840
agç	gcgo	ggg	agga	ataco	gaa d	cccc	tcca	ic c	gato	gtcta	caa	gtag	gtg	ccto	tecat	900
ttt	ttt	acc	atto	caaca	itg a	tgct	gaca	ic ga	ctt	aggg	g gaa	ıttga	ıcgg	tcgt	ggtgc	960
ccc	gago	cctg	acgo	ccag	gat o	cacca	aact	c ga	ıggat	gatt	gga	tcaa	gaa	gcac	gtcag	= 1050
gac	aato	tgg:	tcad	cttcc	atg Met	Ser	aag Lys	Leu	teg Ser	Leu	ago Ser	gag Glu	aag Lys	gag Glu 10	l aaa Lys	1071
gcc Ala	Asn	aac Aen	ttg Leu 15	Pro	gtt Val	cgc Arg	cct Pro	ggc Gly 20	His	ggt Gly	acc Thr	atg Met	ggc Gly 25	Glu	aag Lys	1119
gtg Val	aag Lys	ctt Leu 30	Trp	gcc Ala	aac	tat	ttc Phe 35	aaa Lys	ato	aac Asn	atc Ile	aaa Lys 40	tca Ser	cca Pro	gcc Ala	1167
att Ile	tac Tyr 45	agg Arg	tac	acc Thr	atc Ile	aaa Lys 50	gtt Val	gcc Ala	gcc Ala	acc Thr	gag Glu 55	gaa Glu	'aag Lys	ctc Leu	gga Gly	1215
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ctg Leu	ctc Leu	aag Lys	cag Gln	atc Ile 80	gaa Glu	gcc Ala	aac Asn	gtg Val	aaa Lys 85	tcc Ser	gtg Val	gcg Ala	att Ile	gcc Ala 90	agc Ser	1311
						gtg Val										1359
aac	cac	atc	ttt	gag	ata	aca	taa	acc	gag	cca	ant	tcc	220	a aa	224	1407

Asn	Arg	11e		Glu	Val	. Thr	Trp		: Glu	Pro	Ser	Ser 120		Glm	neA i	
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						Lys					Leu				gat Asp 155	1503
						ttt Phe									gcc Ala	1551
						cat His										1599
						ttt Phe										1647
						tcc Ser 210										1695
						acc Thr										1743
			Val			cct Pro										1791
						atg Met	Asp									1839
	Lys					gac Asp										1887
Ala					Glu	ttg Leu 290				Phe						1935
itt	gtt	tat	aaa :	aaa	tgt	tac	cgc .	acg	ctc	aat	ggc	att	act	aac	cat	1983

11 30		l Ty	r Ly	s Ly:	30!		r Arg	Th:	. Le	31(/ Ile	e Ala	a Ası	1 · Arg 315	
gg Gl	c gad y Asj	ga. Gl	a ago u Aro	320 320	Lys	g caa 3 Glr	a aag n Lys	g gat Asp	ggt Gly 325	Lys	gaa Glu	gto Val	cga Arg	tat Tyr 330	ccg Pro	2031
Pro	c tto D Lei	g tto 2 Phe	c ggg e Gly 335	/ Ile	ccç Pro	ggt Gly	gtc Val	Gln 340	Val	ggc Gly	ggc Gly	ccg Pro	Thr	Ser	tgt Cys	2079
Caq Glr	g tto n Phe	tac Tyr 350	Lev	g cgt Arg	gcg Ala	r Cgạ	gag Glu 355	Thr	aag Lys	gat Asp	ggc Gly	gct Ala 360	gcc	Pro	cct Pro	2127
	act Thr 365	Pro					Asn									2175
	aaa Lys															2223
	gtt Val				Glu											2271
	ctg Leu															2319
	gac Asp															2367
	tct Ser 445									Leu						2415
	acg Thr								lle							2463
	gtc Val							Pro								2511
aag	acg	gta	gag	ccg	cag	gac	ggc (ggg	tgg	ttg .	atg a	aag	ttt	gtc	aag	2559

L	/s :	Ihr	Val	Glu 495	Pro	Gln	Asp	Gly	Gly 500	Trp	Leu	Met	Lys	Phe 505	Val	Lys	
		_	_		_	-	-			-		aca Thr		-	-	-	2607
	s (-	-		-			-		gct Ala 535	-		•		2655
	a G											aac Asn				_	2703
												aaa Lys					2751
	_		-	-		-	-	-				gtg Val		•			2799
												aag Lys					2847
	е Т							-	_	_	-	gcc Ala 615	-	•			2895
	r T											gtc Val			_		2943
												aag Lys	_				2991
			Ala									tat Tyr					3039
		hr .				-	Gly		_		-	tcg Ser	-		•		3087
gto	c g	gc ·	ctg	gtc	tca	acc	atc	gac	caa	cac	ctt	gga	caa	tgg	cct	gca	3135

Va	1 G1 68		u Va	l Se	r Th	f 116		o Gl	n His	s Le	u Gly 695		Tr	p Pro	o Ala	
	t Va					o His					r Met				ttt Phe 715	3183
a c	g ga r As _l	c aa p Ly:	g tte s Phe	2 aaq 2 Lys 72(Thi	g egt : Arg	: ctg Leu	gaa Glu	cta Lev 725	Tr	g cgo Arg	ago Ser	aat Asr	730	gca Ala	3231
Ası	c aad n Asr	c cgo n Aro	agt g Sei 735	Leu	ccc Pro	gag Glu	aat Asn	ato Ile 740	Leu	att Ile	tto Phe	cgc Arg	gat Asp 745	Gly	gtc Val	3279
Sea	gaç Glu	g gga 1 Gly 750	/ Glr	tto Phe	cag Gln	atg Met	gtc Val 755	Ile	aag Lys	gac Asp	gag Glu	cta Leu 760	Pro	ctg	gtt Val	3327
cgo	gcc Ala 765	Ala	tgc Cys	aag Lys	ctg	gtg Val 770	tat Tyr	cca Pro	gct Ala	ggc Gly	aag Lys 775	cta Leu	ccg Pro	egt Arg	att Ile	3375
acg Thr 780	Leu	att	gtc Val	tct Ser	gtc Val 785	aag Lys	cgc Arg	cac His	cag Gln	act Thr 790	cgc Arg	ttc Phe	ttc Phe	cca Pro	ácg Thr 795	3423
gac Asp	ccg Pro	aag Lys	cat His	att Ile 800	cac His	ttc Phe	aag Lys	tcc Ser	aag Lys 805	agc Ser	ccc Pro	aag Lys	gag Glu	ggt Gly 810	act	3471
											tgg Trp					3519
cag Gln	gcg Ala	cac His 830	gcg Ala	tcg Ser	ctc Leu	cag Gln	ggc Gly 835	acg Thr	gcc Ala	cgc Arg	tcg Ser	gct Ala 840	cac His	tac Tyr	aca Thr	3567
gtt Val	ctg Leu 845	gtg Val	gat Asp	gag Glu	att Ile	ttc Phe 850	agg Arg	gcc Ala	gac Asp	tat Tyr	gga Gly 855	aac Asn	aag Lys	gcg Ala	gcc Ala	3615
gac Asp 860	acg Thṛ	ctg Leu	gag Glu	cag Gln	ctg Leu 865	acg Thr	cat His	gac Asp	Met	tgt Cys 870	tat Tyr	ctc Leu	ttt Phe	Gly	cga Arg 875	3663
gcc	acc	aag	gct	gtc	agt	atc	tgc	ccg	cct	gcg	tac	tat .	gcc	gac	ttg	3711

Ala Thr Lys Ala Val Ser Ile Cys Pro Pro Ala Tyr Tyr Ala Asp Leu 880 885 890 gtg tgc gac cgg gcg cgt atc cat cag aag gag ctc ttt gac gcc ctc Val Cys Asp Arg Ala Arg Ile His Gln Lys Glu Leu Phe Asp Ala Leu 895 905 gat gaa aac gat agc gtt aag acc gat gat ttc gca aga tgg ggt aac Asp Glu Asn Asp Ser Val Lys Thr Asp Asp Phe Ala Arg Trp Gly Asn 915 tcc ggg gct gtt cat ccc aac ctt agg aac tcc atg tac tat atc 3852 Ser Gly Ala Val His Pro Asn Leu Arg Asn Ser Met Tyr Tyr Ile 925 930 taggettgte aattgtgtge tggaatgtae tggageatat aagtgaegeg atggaageet 3912 aatcgtctct gaatatggat caaagacagc gtttgctttt tcgggggcttc tagtttctac 3972 agcgatttgt gtggattgtt tettgttetg tttettggtt etttettet ttttttgtg 4032 tetetgtetg cetttgtaet geatgeaaac gtgeaetetg aatgatgaac gacaccattt 4092 gacgattgga taagagatga cagactgcag atactatcat gcgcaatgga aaacacgaac 4152 aaccaaggtt tttgattcct tcaatagcga aatatagaaa aagaaacaaa aaaaaaaaca 4212 acaacaaata atggaagtat gattaaacac attgagcgcg atgactgact ggtgttgtga 4272 atggcgtgtt ggttttcttc tttcttgaaa atttagaacc gtaaatgtta tatcatgtga 4332 tgtaatgtaa taacatattt atatctcgtt gtattcttgt acacactttc caggataaca 4392 tggtctgaca tggtatttct gacgtacaaa aaagaaaaag aaaaacagga aaccatgaac 4452 ccgcgacaaa gctgttccag ttgttacaat gatgatgatg atgatgacct actacctaag 4512 gtattctatc ttagccaagg tattctctcg catcctattc catcctatcc taacccgagc 4572 ctaacccgag cctaaatacc taaactccta aactccttaa ctccttaact cctttctaaa 4632 tgtctaaacc cccaaactat gagacgaccc gaacccgaaa ccctaataaa agtatttata 4692 aaccatcata aaagaaaaaa aaccatcata catggatgat caaaacaaac agaaacggaa 4752 acaacacaac cagctacccg ctcaagactt tcattcgtta attcatcact cactcactca 4812 ctcactcact cagcagcaaa ataccgtttt gtcctgctat tcgtttgttg cgccttgatt 4872

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<210> 2

<211> 938

<212> PRT

<213> Neurospora crassa

<400> 2

Met Ser Lys Leu Ser Leu Ser Glu Lys Glu Lys Ala Asn Asn Leu Pro 1 5 10 15

Val Arg Pro Gly His Gly Thr Met Gly Glu Lys Val Lys Leu Trp Ala 20 25 30

Asn Tyr Phe Lys Ile Asn Ile Lys Ser Pro Ala Ile Tyr Arg Tyr Thr 35 40 45

Ile Lys Val Ala Ala Thr Glu Glu Lys Leu Gly Lys Glu Ala Glu Val

50	55	60
	• • •	ดบ

Ala Ser Lys Lys Val Glu Val Val Val Gly Lys Leu Leu Lys Gln Ile 65 70 75 80

- Glu Ala Asn Val Lys Ser Val Ala Ile Ala Ser Asp Phe Lys Val His 85 90 95
- Leu Val Thr Thr Lys Leu Lys Val Pro Glu Asn Arg Ile Phe Glu 100 105 110
- Val Thr Trp Thr Glu Pro Ser Ser Asn Gln Asn Leu Pro Ser Lys Pro 115 120 125
- Gln Thr Trp Val Val Lys Val Glu Glu Ser Val Glu Thr Cys Asp Phe 130 135 140
- Gly Lys Val Leu Asn Glu Leu Thr Thr Leu Asp Pro Lys Leu Asp Gly 155 160
- Asp Phe Pro Lys Tyr Asn Val Glu Leu Asp Ala Leu Asn Thr Ile Val 165 170 175
- Thr His His Ala Arg Ala Asp Asp Asn Val Ala Val Val Gly Arg Gly 180 185 190
- Arg Phe Phe Ala Ile Gly Asp Asp Leu Ile Glu Gln Val Arg Pro His
 195 200 205
- Asp Ser Pro Leu Val Ile Leu Arg Gly Tyr Phe Ala Ser Val Arg Pro 210 215 220
- Ala Thr Gly Arg Leu Leu Asn Thr Asn Ile Thr His Gly Val Phe 225 230 235 240
- Arg Pro Gly Val Lys Leu Ala Gln Leu Phe Gln Glu Leu Gly Leu Asp
 245 250 255
- Val Met Asp Lys Cys Asn Ala Trp Asn Glu Val Thr Lys Asn Gln Leu 260 265 270
- Asn Asp Lys Met Arg Arg Val His Lys Val Leu Ala Lys Gly Arg Val 275 280 285
- Glu Leu Asn Ala Pro Phe Leu Ile Asp Gly Lys Ile Val Tyr Lys Lys 290 295 300
- Cys Tyr Arg Thr Leu Asn Gly Ile Ala Asn Arg Gly Asp Glu Arg Gly

305	5				310)				315	j				320
Lys	Glr	ı Lys	qeA s	Gly 325		Glu	ı Val	Arg	330		Pro	Leu	Phe	Gly 335	
Pro	Gly	r Va]	340	•	. Gly	Gly	/ Pro	345		: Cys	Glr	Phe	Tyr 350		Arg
Ala	Arg	Glu 355	Thr	Lys	Asp	Gly	7 Ala 360		Pro	Pro	Pro	Thr 365		Gly	Leu
Pro	Ser 370		Ala	Tyr	Ile	Thr 375		Ala	Asn	Tyr	Tyr 380		Gln	Arg	Tyr
Gly 385		Thr	Ala	Asn	Ala 390	Ser	Leu	Pro	Leu	Val 395		Val	Gly	Thr	Lys 400
Glu	Lys	Ala	Ile	Tyr 405	Val	Leu	Ala	Glu	Phe 410		Thr	Leu	Val	Lys 415	Gly
Arg	Ser	Val	Lys 420	Ala	Lys	Leu	Thr	Ala 425	Asn	Glu	Ala	Asp	Asn 430	Met	Ile
Lys	Phe	Ala 435	Суз	Arg	Ala	Pro	Ser 440	Leu	Asn	Ala	Gln	Ser 445	Ile	Val	Thr
Lys	Gly 450	Arg	Gln	Thr	Leu	Gly 455	Leu	Asp	Lys	Ser	Leu 460	Thr	Leu	Gly	Lys
Phe 465	Lys	Val	Ser	Ile	Asp 470	Lys	Glu	Leu	Ile	Thr 475	Val	Val	Gly	Arg	Glu 480
Leu	Lys	Pro	Pro	Met 485	Leu	Thr	Tyr	Ser	Gly 490	Asn	Lys	Thr	Val	Glu 495	Pro
Gln	Asp	Gly	Gly 500	Trp	Leu	Met	Lys	Phe 505	Val	Lys	Val	Ala	Arg 510	Pro	Cys
Arg	Lys	11e 515	Glu	Lys	Trp	Thr	Tyr 520	Leu	Glu	Leu	Lys	Gly 525	Ser	Lys	Ala
Asn	Glu 530	Gly	Val	Pro	Gln	Ala 535	Met	Thr	Ala	Phe	Ala 540	Glu	Phe	Leu	Asn
Arg 545	Thr.	Gly	Ile	Pro	11e 550	Asn	Pro	Arg	Phe	Ser 555	Pro	Gly	Met	Ser	Met 560
Ser	Val	Pro	Gly	Ser	Glu	Lys	Glu	Phe	Phe	Ala	Lys	Val	Lys	Glu	Leu

•	W U U.	1/334	13												PC
			•	565					570					575	
Met	Ser	Ser	His 580	Gln	Phe	Val	Val	Val 585	Leu	Leu	Pro	Arg	Lys 590	Asp	Val
Ala	Ile	Tyr 595	Asn	Met	Val	Lys	Arg 600	Ala	Ala	Asp	Ile	Thr 605	Phe	Gly	Val
His	Thr 610	Val	Cys	Cys	Val	Ala 615	Glu	Lys	Phe	Leu	Ser 620	Thr	Lys	Gly	Gln
Leu 625	Gly	Tyr	Phe	Ala	Asn 630	Val	Gly	Leu	Lys	Val 635	Asn	Leu	Lys	Phe	Gly 640
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Lys	Thr	Met	Val 660	Val	Gly	Tyr	Asp	Val 665	Thr	His	Pro	Thr	Asn 670	Leu	Ala
Ala	Gly	Gln 675	Ser	Pro	Ala	Ser	Ala 680	Pro	Ser	Ile	Val	Gly 685	Leu	Val	Ser
Thr	11e 690	Asp	Gln	His	Leu	Gly 695	Gln	Trp	Pro	Ala	Met 700	Val	Trp	Asn	Asn
Pro 705	His	Gly	Gln	Glu	Ser 710	Met	Thr	Glu	Gln	Phe 715	Thr	Asp	Lys	Phe	Lys 720
Thr	Arg	Leu	Glu	Leu 725	Trp	Arg	Ser	Asn	Pro 730	Ala	Asn	Asn	Arg	Ser 735	Leu
Pro	Glu	Asn	Ile 740	Leu	Ile	Phe	Arg	Asp 745	Gly	Val	Ser	Glu	Gly 750	Gln	Phe
Gln	Met	Val 755	Ile	Lys	Asp	Glu	Leu 760	Pro	Leu	Val	Arg	Ala 765	Ala	Cys	Lys
Leu	Val 770	Tyr	Pro	Ala	Gly	Lys 775	Leu	Pro	Arg	Ile	Thr 780	Leu	Ile	Val	Ser
Val 785	Lys	Arg	His	Gln	Thr 790	Arg	Phe	Phe	Pro	Th i 795	Asp	Pro	Lys	His	Ile 800
His	Phe	Lys	Ser	Lys 805	Ser	Pro	Lys	Glu	Gly 810	Thr	Val	Val	Asp	Arg 815	Gly

Val Thr Asn Val Arg Tyr Trp Asp Phe Phe Leu Gln Ala His Ala Ser

820 825 830

Leu Gln Gly Thr Ala Arg Ser Ala His Tyr Thr Val Leu Val Asp Glu 835 840 845

- Ile Phe Arg Ala Asp Tyr Gly Asn Lys Ala Ala Asp Thr Leu Glu Gln 850 855 860
- Leu Thr His Asp Met Cys Tyr Leu Phe Gly Arg Ala Thr Lys Ala Val 865 870 870 875 880
- Ser Ile Cys Pro Pro Ala Tyr Tyr Ala Asp Leu Val Cys Asp Arg Ala 885 890 895
- Arg Ile His Gln Lys Glu Leu Phe Asp Ala Leu Asp Glu Asn Asp Ser 900 . 905 910
- Val Lys Thr Asp Asp Phe Ala Arg Trp Gly Asn Ser Gly Ala Val His 915 920 925
- Pro Asn Leu Arg Asn Ser Met Tyr Tyr Ile 930 935